



# Relationship of Aberrant DNA Methylation in Inheritable and Sporadic Endometrial Cancer Carcinogenesis

著者	MOROYAMA Megumi
内容記述	この博士論文は内容の要約のみの公開（または一部非公開）になっています
year	2017
その他のタイトル	遺伝性・散発性子宮体癌の発癌におけるDNA異常メチル化の関わり
学位授与大学	筑波大学 (University of Tsukuba)
学位授与年度	2017
報告番号	12102甲第8330号
URL	<a href="http://hdl.handle.net/2241/00150048">http://hdl.handle.net/2241/00150048</a>

**Relationship of Aberrant DNA Methylation  
in Inheritable and Sporadic  
Endometrial Cancer Carcinogenesis**

**May 2017**

**Megumi MOROYAMA**



**Relationship of Aberrant DNA Methylation  
in Inheritable and Sporadic  
Endometrial Cancer Carcinogenesis**

A Dissertation Submitted to the Graduate School of Life and Environmental  
Sciences, the University of Tsukuba in Partial Fulfillment of the  
Requirements for the Degree of Doctor of Philosophy in Science  
(Doctoral Program in Functional Biological Sciences)

**Megumi MOROYAMA**



# Contents

Abstract .....	1
Abbreviations .....	4
General Introduction .....	6
General Discussion .....	14
References .....	18
Acknowledgements .....	45

## **Abstract**

DNA methylation is essential for healthy mammalian development and function, and is involved in important processes such as suppression of repetitive elements, genomic imprinting and carcinogenesis. Thus, aberrant DNA methylation is linked to some of the adverse outcomes. DNA methylation indicates addition of a methyl ( $\text{CH}_3$ ) group to fifth position of a cytosine within CpG dinucleotides, which forms clusters called CpG islands. DNA methylation patterns are established and maintained by DNA methyltransferases (DNMTs). Inactivation of certain tumor suppressor genes caused by methylation of the promoter region is widely observed in various types of cancer.

Epigenetic reprogramming, including DNA demethylation, occurs in mammalian primordial germ cells (PGCs) and early embryos, and returns the cells to pluripotency. Therefore, DNA methylation in human cells has long been thought not to be inherited. However, recent studies have raised the possibility that epimutation can occur in human cells.

In chapter 1, I focus on Lynch syndrome, an inherited cancer syndrome that is caused by germline mutation of DNA mismatch repair (MMR) genes and has an increased risk of colorectal, endometrial and other cancers. Recent studies have shown that 25-30% of patients with Lynch syndrome have no germline mutation of MMR genes. This raises the possibility that epimutation of MMR genes could be an alternative cause of Lynch syndrome. Therefore, I investigated epimutation of MMR genes in peripheral blood DNA in 106 patients with endometrial cancer, and identified patients with Lynch syndrome.

Epimutation could be a cause of inherited cancer syndrome and sporadic cancer. Some types of cancers show concomitant DNA methylation of multiple genes, which is referred to as CpG island methylator phenotype (CIMP). In colon cancer, Lynch syndrome and CIMP-positive cancer show similar clinicopathological features. However, there are few studies on CIMP-positive endometrial cancer, and the causes and features of this condition are unknown. In chapter 2, I hypothesized that patients with



CIMP-positive endometrial cancer have aberrant DNA methylation (epimutation) in normal tissue that is a trigger of carcinogenesis. Therefore, I investigated the genome-wide methylation status of DNA from peripheral blood cells (PBCs) and cancer tissue in patients with CIMP-positive and CIMP-negative endometrial cancer. In DNA from PBCs, the promoter region of miR-663a was significantly hypermethylated in CIMP-positive cases compared to CIMP-negative cases.

These studies provide new knowledge on the relationship between epigenetic abnormalities and endometrial carcinogenesis. The findings may be applicable to early detection and as a predictive marker. Furthermore, DNA methylation is reversible, and future strategies for DNA demethylation may contribute to cancer prevention.

## **Abbreviations**

Atypical endometrial hyperplasia: AEH

CIMP-high: CIMP-H

CIMP-low: CIMP-L

CIMP-negative: CIMP(-)

CpG island methylator phenotype: CIMP

Differentially methylated CpGs: DMCs

Differentially methylated regions: DMRs

DNA methyltransferase: DNMT

DNA mismatch repair gene: MMR gene

Methylation specific polymerase chain reaction: MSP

MicroRNAs: miRNAs

Next-generation sequencing: NGS

Peripheral blood cells: PBCs

Polymerase chain reaction: PCR

Post-Bisulfite Adaptor Tagging: PBAT

Primordial germ cells: PGCs

Transcription start sites: TSS

## **General Introduction**

DNA methylation is one of the epigenetic mechanisms used to regulate gene expression. Among several mechanisms regulating gene expression, DNA methylation is the most common for fixing genes in the “off” position. Thus, DNA methylation plays important roles in embryonic development, chromosome stability and carcinogenesis. Indeed, the relationship between methylation abnormalities and human diseases such as cancer, psychiatric disorder and congenital imprinting disorders are currently being studied (1). The results of these studies will be important for not only treatment of these diseases but also understanding of DNA methylation mechanisms and prevention of DNA methylation abnormalities.

DNA methylation occurs by the addition of methyl groups to cytosine bases in mammalian DNA by DNMTs. In mammals, there are 3 major DNMTs: DNMT1, DNMT3a and DNMT3b. DNMT3a and DNMT3b are de novo DNMTs that show equal affinity for hemi-methylated DNA (DNA with one strand methylated) and non-methylated DNA (2). In contrast,

DNMT1 is a maintenance DNMT that binds to hemi-methylated DNA at CpG sites. After DNA replication, the parent strand remains methylated, but the daughter strand is not methylated. DNMT1 binds to these hemi-methylated CpGs and methylates the cytosine on the newly synthesized daughter strand, and maintains CpG methylation patterns through mitosis (3).

Unlike animals, plants do not have a separate germline in which epigenetic marks are erased and reestablished. Thus, even if DNA methylation machinery is restored, epigenetic changes induced in DNA methylation abnormalities can be maintained and inherited (4). One of the oldest examples of heritable epigenetic change (epimutation) in plants is a morphological defect in the development of flower in *Linaria vulgaris*. The mutant phenotype is due to aberrant DNA methylation and transcriptional suppression of *Lcyc*, which is a regulator of dorsoventral asymmetry (Figure 1). In correlation with the expression recovery by demethylation of *Lcyc* gene, phenotype is restored occasionally (5).

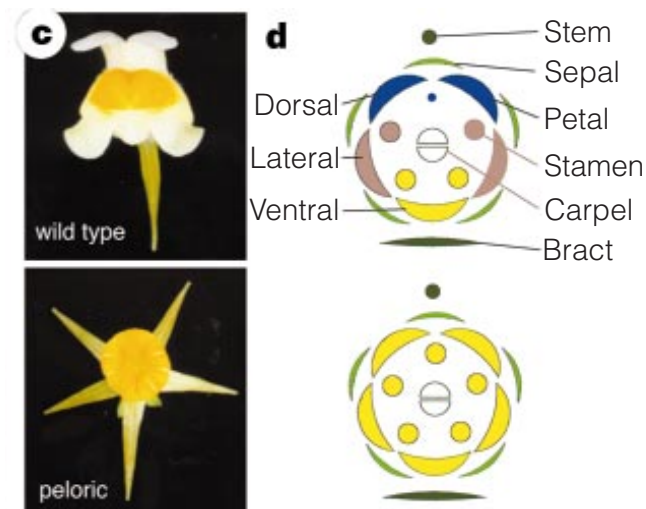


Figure 1 Wild-type and peloric *Linaria vulgaris* flowers.

c, Face view of a wild-type *Linaria* flower compared to a peloric mutant.

d, Floral diagrams of wild-type (top) and peloric (bottom) flowers showing the relative positions of different organs.

Adapted from Cubas, P. et al. :Nature, 401: 157-161, 1999

In contrast, methylation patterns of mammalian cells are erased in PGCs and at the post-fertilization stage (Figure 2). Therefore, epimutation was thought not to occur in mammals. However, there is increasing evidence that environmental (nutritional) stimuli can modify DNA methylation and affect phenotypic expression of genes. For example, the methylation level of the *Leptin* promoter is significantly increased in oocytes of high-fat diet mice. Female offspring from the obese mice showed higher methylation level of

*Leptin* promoter in the liver than normal mice. Expression level of *Leptin* was also significantly decreased in the liver of these offspring (6). Examples of epigenetic inheritance in mice raise possibility that occurs similar event in humans.

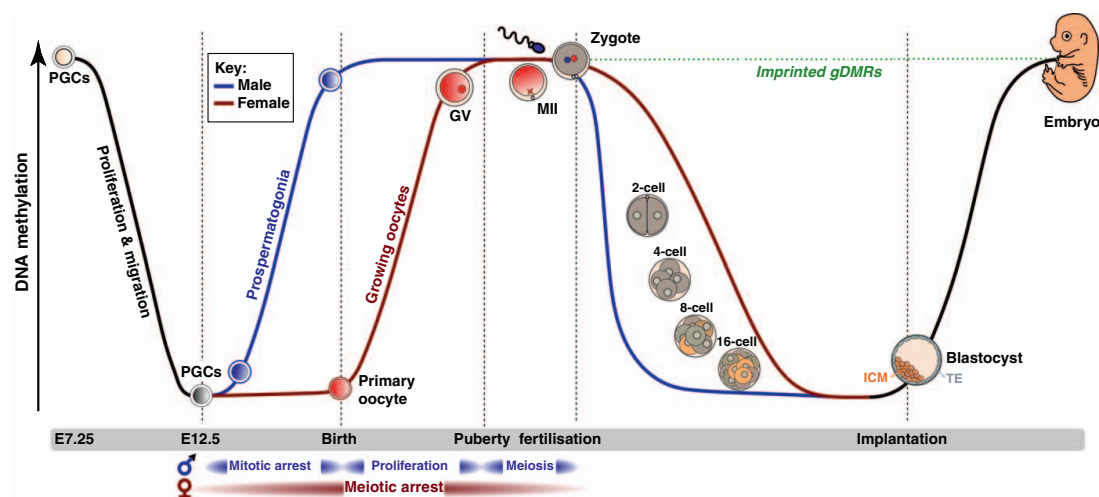


Figure 2. DNA methylation changes during developmental epigenetic reprogramming.

Adapted from Smallwood, SA. et al. : *Trans Genet*, 28: 33-42, 2012

To examine this issue further, I focused on Lynch syndrome, which is an inherited cancer syndrome caused by germline mutation of DNA

mismatch repair (MMR) genes, most frequently *MLH1* and *MSH2* (7, 8).

Lynch syndrome is characterized by increased risk of colorectal, endometrial, ovarian and other cancers. In Knudson's two-hit theory, it is required that an abnormality (hit) occurs in both alleles of a tumor-suppressor gene for disease progression (Figure 3). Germline mutations generally represent the

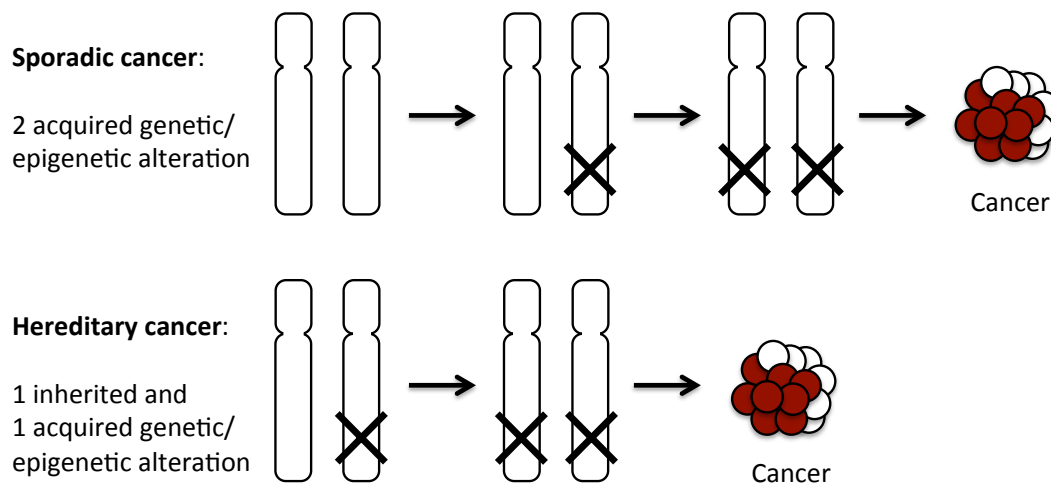


Figure 3. Knudson's two-hit theory

Modified from [http://ocw.tufts.edu/data/20/300759/300840\\_xlarge.jpg](http://ocw.tufts.edu/data/20/300759/300840_xlarge.jpg)

first hit on one allele, while the second hit typically results from a sporadic mutation, loss of heterozygosity or methylation. However, germline



mutations of MMR genes are not found in 25-30% of patients with Lynch syndrome (9-12). Therefore, it is possible that epimutation of MMR genes acts as the first hit in patients with no germline mutation.

Evaluation of epimutation of MMR genes to date has been based on case studies with insufficient information on families, and epimutation in families with endometrial cancer has not been examined. In chapter 1, I explored epimutation in cases of endometrial cancer and identified patients with Lynch syndrome.

Epimutation could be a cause of inherited cancer syndrome and sporadic cancer. Aberrant DNA hypermethylation in CpG island is a hallmark of cancer and is characterized by tumor-specific hypermethylation of numerous CpG islands (13). *MLH1* methylation is also observed in cases of sporadic colorectal and endometrial cancer (14). These cancers show the same phenotype of mismatch repair defect and clinicopathologic characteristics similar to Lynch syndrome. Such sporadic colorectal cancer also has a close relationship with cancer with a CpG island methylator

phenotype (CIMP) (15) (Figure 4). CIMP was first proposed by Toyota et al.

in 1999 (16). They defined a subgroup of colorectal cancers with concurrent

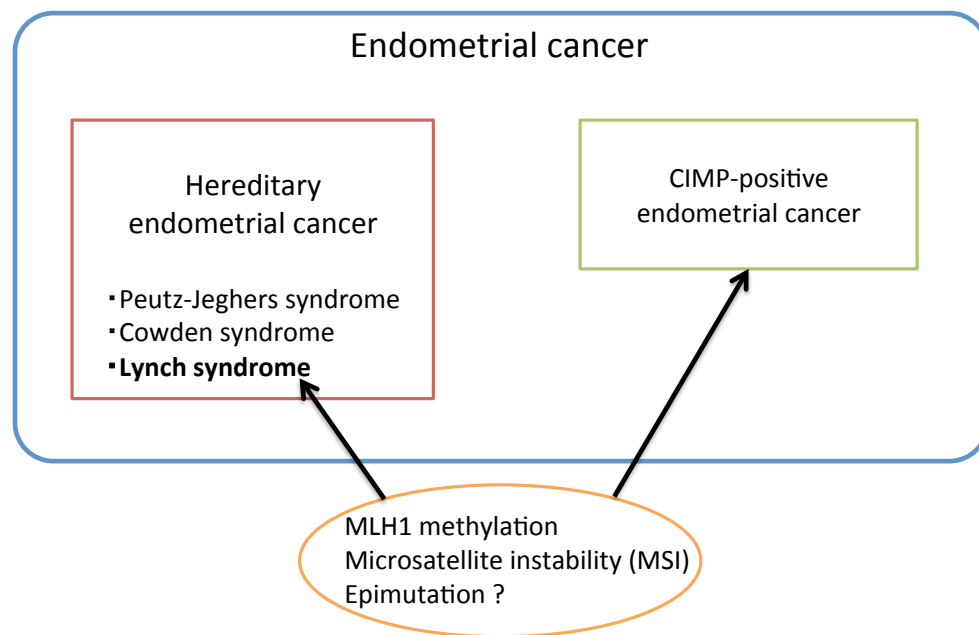


Figure 4. Relationship between Lynch syndrome and CIMP-positive endometrial cancer

multiple promoter hypermethylation of tumor-related genes as CIMP.

CIMP has subsequently been reported in gastric (17, 18), lung (19, 20), liver

(21) and ovarian (22) cancer. Recent studies revealed that CIMP is

negatively associated with genetic mutations in colorectal cancers (23, 24),

which suggests that it can provide an alternative oncogenic pathway. This findings supported that epimutation may be one of the cause of CIMP positive cancer. Thus, I hypothesized that normal tissue of CIMP-positive endometrial cancer has features which are prone to get DNA methylation, and that such features play important roles in carcinogenesis in this cancer.

## General Discussion

Lynch syndrome is one of the most prevalent hereditary cancer syndromes in humans and is caused by inherited defects in MMR genes (89, 90). In the last two decades, increased appreciation of epigenetic mechanisms in tumorigenesis and identification of constitutional epimutations underlying Lynch syndrome have laid the foundation for the epigenetic era (11). Epimutation is regarded as secondary if induced by an adjacent genetic alteration, and otherwise as primary (33). Lynch syndrome offers one of the first examples of cancer-associated constitutional epimutation, namely primary epimutation of *MLH1* (11). Recent observations of constitutional epimutations as the first hit and promoter methylation as the second hit in Lynch syndrome emphasize the increasing significance of epigenetic events, especially as methylation as the second hit is associated with a more generalized CIMP in tumors (91).

CIMP was first explained by Toyota et al. in 1999 (16). CIMP occurs in a subset of colorectal cancers that are characterized by vast

hypermethylation of promoter CpG island sites, resulting in inactivation of several tumor suppressor genes or other tumor-related genes (92). Many studies have found an association between CIMP status and other important epidemiological and molecular factors, such as smoking, age and genetic mutations (93-95).

In chapter 1, I tried to identify MMR genes epimutation positive patients from 106 endometrial cancer patients. According to our preliminary experiment, 1% of endometrial cancer patients show *MLH1* epimutation, but no cases of epimutation-positive endometrial cancer were found in this study.

I also identified two patients with Lynch syndrome among the 106 patients with endometrial cancer, based on the Amsterdam II criteria and revised Bethesda guidelines (diagnostic criteria for Lynch syndrome using family and personal history). One of these patients had a novel *MSH6* nonsense mutation. Since colon cancer in patients with Lynch syndrome is characterized by mutations in *MLH1* and *MSH2*, rather than *MSH6*,

endometrial cancer with Lynch syndrome may have a different carcinogenetic pathway.

Next, I focused on the relationship between epigenetic alteration and sporadic endometrial carcinogenesis. Many studies have shown that DNA methylation is related to endometrial cancer, especially in the early stages of carcinogenesis. Moreover, some types of cancer, including endometrial cancer, show a CIMP phenotype. These findings suggest that DNA methylation plays important roles in carcinogenesis and in cancer phenotypes, but it is still unclear whether such DNA methylation is a cause or a result of cancer.

I succeeded in identifying aberrant DNA methylation, which is a potential cause of CIMP-positive endometrial carcinogenesis. The MiR-663a promoter region in PBC DNA of CIMP-H patients was significantly hypermethylated compared to that of CIMP(-) patients. Consistent with this methylation pattern, miR-663a expression in PBCs of CIMP-H patients was lower than that in CIMP(-) patients. The miR-663a promoter region is a

hypomethylated region in normal human adult tissues, and therefore miR-663a DNA methylation identified in this study may be a novel epimutation candidate.

This study provides new findings on the involvement of epigenetic abnormalities in hereditary and sporadic endometrial cancer. Detection of abnormal DNA methylation using a blood specimen can be performed quickly and conveniently, and thus is useful as a cancer prediction and diagnostic marker. Furthermore, since methylation of DNA is reversible, site-specific demethylation may contribute to prevention of cancer.

## References

1. Robertson KD: DNA methylation and human disease. *Nature reviews. Genetics* 6: 597-610, 2005.
2. Okano M, Xie S and Li E: Cloning and characterization of a family of novel mammalian DNA (cytosine-5) methyltransferases. *Nature genetics* 19: 219-220, 1998.
3. Bestor TH: The DNA methyltransferases of mammals. *Human molecular genetics* 9: 2395-2402, 2000.
4. Reinders J, Wulff BB, Mirouze M, Mari-Ordonez A, Dapp M, Rozhon W, Bucher E, Theiler G and Paszkowski J: Compromised stability of DNA methylation and transposon immobilization in mosaic *Arabidopsis* epigenomes. *Genes & development* 23: 939-950, 2009.
5. Cubas P, Vincent C and Coen E: An epigenetic mutation responsible for natural variation in floral symmetry. *Nature* 401: 157-161, 1999.
6. Ge ZJ, Luo SM, Lin F, Liang QX, Huang L, Wei YC, Hou Y, Han ZM,



Schatten H and Sun QY: DNA methylation in oocytes and liver of female mice and their offspring: effects of high-fat-diet-induced obesity. *Environmental health perspectives* 122: 159-164, 2014.

7. Banno K, Susumu N, Hirao T, Yanokura M, Hirasawa A, Aoki D, Udagawa Y, Sugano K and Nozawa S: Identification of germline MSH2 gene mutations in endometrial cancer not fulfilling the new clinical criteria for hereditary nonpolyposis colorectal cancer. *Cancer genetics and cytogenetics* 146: 58-65, 2003.

8. Quehenberger F, Vasen HF and van Houwelingen HC: Risk of colorectal and endometrial cancer for carriers of mutations of the hMLH1 and hMSH2 gene: correction for ascertainment. *Journal of medical genetics* 42: 491-496, 2005.

9. Chan TL, Yuen ST, Kong CK, Chan YW, Chan AS, Ng WF, Tsui WY, Lo MW, Tam WY, Li VS and Leung SY: Heritable germline epimutation of MSH2 in a family with hereditary nonpolyposis colorectal cancer. *Nature genetics* 38: 1178-1183, 2006.

10. Cini G, Carnevali I, Quaia M, Chiaravalli AM, Sala P, Giacomini E, Maestro R, Tibiletti MG and Viel A: Concomitant mutation and epimutation of the MLH1 gene in a Lynch syndrome family. *Carcinogenesis* 36: 452-458, 2015.
11. Gazzoli I, Loda M, Garber J, Syngal S and Kolodner RD: A hereditary nonpolyposis colorectal carcinoma case associated with hypermethylation of the MLH1 gene in normal tissue and loss of heterozygosity of the unmethylated allele in the resulting microsatellite instability-high tumor. *Cancer research* 62: 3925-3928, 2002.
12. Suter CM, Martin DI and Ward RL: Germline epimutation of MLH1 in individuals with multiple cancers. *Nature genetics* 36: 497-501, 2004.
13. Costello JF, Fruhwald MC, Smiraglia DJ, Rush LJ, Robertson GP, Gao X, Wright FA, Feramisco JD, Peltomaki P, Lang JC, Schuller DE, Yu L, Bloomfield CD, Caligiuri MA, Yates A, Nishikawa R, Su Huang H, Petrelli NJ, Zhang X, O'Dorisio MS, Held WA, Cavenee WK and Plass C: Aberrant CpG-island methylation has non-random and tumour-type-specific patterns.

Nature genetics 24: 132-138, 2000.

14. Kane MF, Loda M, Gaida GM, Lipman J, Mishra R, Goldman H, Jessup JM and Kolodner R: Methylation of the hMLH1 promoter correlates with lack of expression of hMLH1 in sporadic colon tumors and mismatch repair-defective human tumor cell lines. Cancer research 57: 808-811, 1997.

15. Weisenberger DJ, Siegmund KD, Campan M, Young J, Long TI, Faasse MA, Kang GH, Widschwendter M, Weener D, Buchanan D, Koh H, Simms L, Barker M, Leggett B, Levine J, Kim M, French AJ, Thibodeau SN, Jass J, Haile R and Laird PW: CpG island methylator phenotype underlies sporadic microsatellite instability and is tightly associated with BRAF mutation in colorectal cancer. Nature genetics 38: 787-793, 2006.

16. Toyota M, Ahuja N, Ohe-Toyota M, Herman JG, Baylin SB and Issa JP: CpG island methylator phenotype in colorectal cancer. Proceedings of the National Academy of Sciences of the United States of America 96: 8681-8686, 1999.

17. Loh M, Liem N, Vaithilingam A, Lim PL, Sapari NS, Elahi E, Mok

ZY, Cheng CL, Yan B, Pang B, Salto-Tellez M, Yong WP, Iacopetta B and Soong R: DNA methylation subgroups and the CpG island methylator phenotype in gastric cancer: a comprehensive profiling approach. *BMC gastroenterology* 14: 55, 2014.

18. Shigeyasu K, Nagasaka T, Mori Y, Yokomichi N, Kawai T, Fuji T, Kimura K, Umeda Y, Kagawa S, Goel A and Fujiwara T: Clinical Significance of MLH1 Methylation and CpG Island Methylator Phenotype as Prognostic Markers in Patients with Gastric Cancer. *PloS one* 10: e0130409, 2015.

19. Saito Y, Nagae G, Motoi N, Miyauchi E, Ninomiya H, Uehara H, Mun M, Okumura S, Ohyanagi F, Nishio M, Satoh Y, Aburatani H and Ishikawa Y: Prognostic significance of CpG island methylator phenotype in surgically resected small cell lung carcinoma. *Cancer science* 107: 320-325, 2016.

20. Shinjo K, Okamoto Y, An B, Yokoyama T, Takeuchi I, Fujii M, Osada H, Usami N, Hasegawa Y, Ito H, Hida T, Fujimoto N, Kishimoto T,

Sekido Y and Kondo Y: Integrated analysis of genetic and epigenetic alterations reveals CpG island methylator phenotype associated with distinct clinical characters of lung adenocarcinoma. *Carcinogenesis* 33: 1277-1285, 2012.

21. Cheng Y, Zhang C, Zhao J, Wang C, Xu Y, Han Z, Jiang G, Guo X, Li R, Bu X, Wu M and Wei L: Correlation of CpG island methylator phenotype with poor prognosis in hepatocellular carcinoma. *Experimental and molecular pathology* 88: 112-117, 2010.

22. Strathdee G, Appleton K, Illand M, Millan DW, Sargent J, Paul J and Brown R: Primary ovarian carcinomas display multiple methylator phenotypes involving known tumor suppressor genes. *The American journal of pathology* 158: 1121-1127, 2001.

23. Goel A, Nagasaka T, Arnold CN, Inoue T, Hamilton C, Niedzwiecki D, Compton C, Mayer RJ, Goldberg R, Bertagnolli MM and Boland CR: The CpG island methylator phenotype and chromosomal instability are inversely correlated in sporadic colorectal cancer. *Gastroenterology* 132: 127-138,

2007.

24. Ogino S, Kawasaki T, Kirkner GJ, Ohnishi M and Fuchs CS: 18q loss of heterozygosity in microsatellite stable colorectal cancer is correlated with CpG island methylator phenotype-negative (CIMP-0) and inversely with CIMP-low and CIMP-high. *BMC cancer* 7: 72, 2007.

25. Lynch HT: Hereditary nonpolyposis colorectal cancer (HNPCC). *Cytogenetics and cell genetics* 86: 130-135, 1999.

26. Peltomaki P: Role of DNA mismatch repair defects in the pathogenesis of human cancer. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 21: 1174-1179, 2003.

27. Banno K, Yanokura M, Iida M, Masuda K and Aoki D: Carcinogenic mechanisms of endometrial cancer: involvement of genetics and epigenetics. *The journal of obstetrics and gynaecology research* 40: 1957-1967, 2014.

28. Banno K, Kisu I, Yanokura M, Masuda K, Ueki A, Kobayashi Y, Susumu N and Aoki D: Epigenetics and genetics in endometrial cancer: new carcinogenic mechanisms and relationship with clinical practice.

Epigenomics 4: 147-162, 2012.

29. Banno K, Kisu I, Yanokura M, Tsuji K, Masuda K, Ueki A, Kobayashi Y, Yamagami W, Nomura H, Tominaga E, Susumu N and Aoki D: Epimutation and cancer: a new carcinogenic mechanism of Lynch syndrome (Review). International journal of oncology 41: 793-797, 2012.

30. Masuda K, Banno K, Hirasawa A, Yanokura M, Tsuji K, Kobayashi Y, Kisu I, Ueki A, Nomura H, Tominaga E, Susumu N and Aoki D: Relationship of lower uterine segment cancer with Lynch syndrome: a novel case with an hMLH1 germline mutation. Oncology reports 28: 1537-1543, 2012.

31. Masuda K, Banno K, Yanokura M, Kobayashi Y, Kisu I, Ueki A, Ono A, Asahara N, Nomura H, Hirasawa A, Susumu N and Aoki D: Relationship between DNA Mismatch Repair Deficiency and Endometrial Cancer. Molecular biology international 2011: 256063, 2011.

32. Hitchins MP: Finding the needle in a haystack: identification of cases of Lynch syndrome with MLH1 epimutation. Familial cancer 15:

413-422, 2016.

33. Hitchins MP: Constitutional epimutation as a mechanism for cancer causality and heritability? *Nature reviews. Cancer* 15: 625-634, 2015.

34. Ward RL, Dobbins T, Lindor NM, Rapkins RW and Hitchins MP: Identification of constitutional MLH1 epimutations and promoter variants in colorectal cancer patients from the Colon Cancer Family Registry. *Genetics in medicine : official journal of the American College of Medical Genetics* 15: 25-35, 2013.

35. Castillejo A, Hernandez-Illan E, Rodriguez-Soler M, Perez-Carbonell L, Egoavil C, Barbera VM, Castillejo MI, Guarinos C, Martinez-de-Duenas E, Juan MJ, Sanchez-Heras AB, Garcia-Casado Z, Ruiz-Ponte C, Brea-Fernandez A, Juarez M, Bujanda L, Clofent J, Llor X, Andreu M, Castells A, Carracedo A, Alenda C, Paya A, Jover R and Soto JL: Prevalence of MLH1 constitutional epimutations as a cause of Lynch syndrome in unselected versus selected consecutive series of patients with colorectal cancer. *Journal of medical genetics* 52: 498-502, 2015.



36. Morak M, Schackert HK, Rahner N, Betz B, Ebert M, Walldorf C, Royer-Pokora B, Schulmann K, von Knebel-Doeberitz M, Dietmaier W, Keller G, Kerker B, Leitner G and Holinski-Feder E: Further evidence for heritability of an epimutation in one of 12 cases with MLH1 promoter methylation in blood cells clinically displaying HNPCC. *European journal of human genetics : EJHG* 16: 804-811, 2008.
37. Pineda M, Mur P, Iniesta MD, Borrás E, Campos O, Vargas G, Iglesias S, Fernandez A, Gruber SB, Lazaro C, Brunet J, Navarro M, Blanco I and Capella G: MLH1 methylation screening is effective in identifying epimutation carriers. *European journal of human genetics : EJHG* 20: 1256-1264, 2012.
38. Miyaki M, Konishi M, Tanaka K, Kikuchi-Yanoshita R, Muraoka M, Yasuno M, Igari T, Koike M, Chiba M and Mori T: Germline mutation of MSH6 as the cause of hereditary nonpolyposis colorectal cancer. *Nature genetics* 17: 271-272, 1997.
39. Wijnen J, de Leeuw W, Vasen H, van der Klift H, Moller P,

Stormorken A, Meijers-Heijboer H, Lindhout D, Menko F, Vossen S, Moslein G, Tops C, Brocker-Vriends A, Wu Y, Hofstra R, Sijmons R, Cornelisse C, Morreau H and Fodde R: Familial endometrial cancer in female carriers of MSH6 germline mutations. *Nature genetics* 23: 142-144, 1999.

40. Hirai Y, Banno K, Suzuki M, Ichikawa Y, Udagawa Y, Sugano K and Miki Y: Molecular epidemiological and mutational analysis of DNA mismatch repair (MMR) genes in endometrial cancer patients with HNPCC-associated familial predisposition to cancer. *Cancer science* 99: 1715-1719, 2008.

41. Banno K, Yanokura M, Kobayashi Y, Kawaguchi M, Nomura H, Hirasawa A, Susumu N and Aoki D: Endometrial cancer as a familial tumor: pathology and molecular carcinogenesis (review). *Current genomics* 10: 127-132, 2009.

42. Fiolka R, Zubor P, Janusicova V, Visnovsky J, Mendelova A, Kajo K, Lasabova Z, Plank L and Danko J: Promoter hypermethylation of the tumor-suppressor genes RASSF1A, GSTP1 and CDH1 in endometrial cancer.

Oncology reports 30: 2878-2886, 2013.

43. Guida M, Sanguedolce F, Bufo P, Di Spiezio Sardo A, Bifulco G, Nappi C and Pannone G: Aberrant DNA hypermethylation of hMLH-1 and CDKN2A/p16 genes in benign, premalignant and malignant endometrial lesions. European journal of gynaecological oncology 30: 267-270, 2009.

44. Kanaya T, Kyo S, Maida Y, Yatabe N, Tanaka M, Nakamura M and Inoue M: Frequent hypermethylation of MLH1 promoter in normal endometrium of patients with endometrial cancers. Oncogene 22: 2352-2360, 2003.

45. Crepin M, Dieu MC, Lejeune S, Escande F, Boidin D, Porchet N, Morin G, Manouvrier S, Mathieu M and Buisine MP: Evidence of constitutional MLH1 epimutation associated to transgenerational inheritance of cancer susceptibility. Human mutation 33: 180-188, 2012.

46. Romero-Perez L, Lopez-Garcia MA, Diaz-Martin J, Biscuola M, Castilla MA, Tafe LJ, Garg K, Oliva E, Matias-Guiu X, Soslow RA and Palacios J: ZEB1 overexpression associated with E-cadherin and

microRNA-200 downregulation is characteristic of undifferentiated endometrial carcinoma. *Modern pathology : an official journal of the United States and Canadian Academy of Pathology, Inc* 26: 1514-1524, 2013.

47. Bischoff J, Ignatov A, Semczuk A, Schwarzenau C, Ignatov T, Krebs T, Kuster D, Przada-Rabaniuk D, Roessner A, Costa SD and Schneider-Stock R: hMLH1 promoter hypermethylation and MSI status in human endometrial carcinomas with and without metastases. *Clinical & experimental metastasis* 29: 889-900, 2012.

48. Weisenberger DJ, Levine AJ, Long TI, Buchanan DD, Walters R, Clendenning M, Rosty C, Joshi AD, Stern MC, Le Marchand L, Lindor NM, Daftary D, Gallinger S, Selander T, Bapat B, Newcomb PA, Campbell PT, Casey G, Ahnen DJ, Baron JA, Haile RW, Hopper JL, Young JP, Laird PW and Siegmund KD: Association of the colorectal CpG island methylator phenotype with molecular features, risk factors, and family history. *Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of*

Preventive Oncology 24: 512-519, 2015.

49. Zhang QY, Yi DQ, Zhou L, Zhang DH and Zhou TM: Status and significance of CpG island methylator phenotype in endometrial cancer.

Gynecologic and obstetric investigation 72: 183-191, 2011.

50. Park SY, Kook MC, Kim YW, Cho NY, Jung N, Kwon HJ, Kim TY and Kang GH: CpG island hypermethylator phenotype in gastric carcinoma and its clinicopathological features. Virchows Archiv : an international journal of pathology 457: 415-422, 2010.

51. Samowitz WS, Albertsen H, Herrick J, Levin TR, Sweeney C, Murtaugh MA, Wolff RK and Slattery ML: Evaluation of a large, population-based sample supports a CpG island methylator phenotype in colon cancer. Gastroenterology 129: 837-845, 2005.

52. Whitcomb BP, Mutch DG, Herzog TJ, Rader JS, Gibb RK and Goodfellow PJ: Frequent HOXA11 and THBS2 promoter methylation, and a methylator phenotype in endometrial adenocarcinoma. Clinical cancer research : an official journal of the American Association for Cancer

Research 9: 2277-2287, 2003.

53. Peterson LM, Kipp BR, Halling KC, Kerr SE, Smith DI, Distad TJ, Clayton AC and Medeiros F: Molecular characterization of endometrial cancer: a correlative study assessing microsatellite instability, MLH1 hypermethylation, DNA mismatch repair protein expression, and PTEN, PIK3CA, KRAS, and BRAF mutation analysis. *International journal of gynecological pathology : official journal of the International Society of Gynecological Pathologists* 31: 195-205, 2012.

54. Kawaguchi M, Yanokura M, Banno K, Kobayashi Y, Kuwabara Y, Kobayashi M, Nomura H, Hirasawa A, Susumu N and Aoki D: Analysis of a correlation between the BRAF V600E mutation and abnormal DNA mismatch repair in patients with sporadic endometrial cancer. *International journal of oncology* 34: 1541-1547, 2009.

55. Banno K, Kisu I, Yanokura M, Masuda K, Kobayashi Y, Ueki A, Tsuji K, Yamagami W, Nomura H, Susumu N and Aoki D: Endometrial Cancer and Hypermethylation: Regulation of DNA and MicroRNA by

Epigenetics. Biochemistry research international 2012: 738274, 2012.

56. Yanokura M, Banno K, Kawaguchi M, Hirao N, Hirasawa A, Susumu N, Tsukazaki K and Aoki D: Relationship of aberrant DNA hypermethylation of CHFR with sensitivity to taxanes in endometrial cancer. Oncology reports 17: 41-48, 2007.

57. Yanokura M, Banno K, Susumu N, Kawaguchi M, Kuwabara Y, Tsukazaki K and Aoki D: Hypermethylation in the p16 promoter region in the carcinogenesis of endometrial cancer in Japanese patients. Anticancer research 26: 851-856, 2006.

58. Banno K, Yanokura M, Susumu N, Kawaguchi M, Hirao N, Hirasawa A, Tsukazaki K and Aoki D: Relationship of the aberrant DNA hypermethylation of cancer-related genes with carcinogenesis of endometrial cancer. Oncology reports 16: 1189-1196, 2006.

59. Akalin A, Kormaksson M, Li S, Garrett-Bakelman FE, Figueroa ME, Melnick A and Mason CE: methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation profiles. Genome biology 13: R87,

2012.

60. Avraham A, Cho SS, Uhlmann R, Polak ML, Sandbank J, Karni T, Pappo I, Halperin R, Vaknin Z, Sella A, Sukumar S and Evron E: Tissue specific DNA methylation in normal human breast epithelium and in breast cancer. *PloS one* 9: e91805, 2014.

61. Miura F, Enomoto Y, Dairiki R and Ito T: Amplification-free whole-genome bisulfite sequencing by post-bisulfite adaptor tagging. *Nucleic acids research* 40: e136, 2012.

62. Alvarez H, Opalinska J, Zhou L, Sohal D, Fazzari MJ, Yu Y, Montagna C, Montgomery EA, Canto M, Dunbar KB, Wang J, Roa JC, Mo Y, Bhagat T, Ramesh KH, Cannizzaro L, Mollenhauer J, Thompson RF, Suzuki M, Meltzer SJ, Melnick A, Greally JM, Maitra A and Verma A: Widespread hypomethylation occurs early and synergizes with gene amplification during esophageal carcinogenesis. *PLoS genetics* 7: e1001356, 2011.

63. Feinberg AP and Vogelstein B: Hypomethylation distinguishes genes of some human cancers from their normal counterparts. *Nature* 301:



89-92, 1983.

64. Kundaje A, Meuleman W, Ernst J, Bilenky M, Yen A, Heravi-Moussavi A, Kheradpour P, Zhang Z, Wang J, Ziller MJ, Amin V, Whitaker JW, Schultz MD, Ward LD, Sarkar A, Quon G, Sandstrom RS, Eaton ML, Wu YC, Pfenning AR, Wang X, Claussnitzer M, Liu Y, Coarfa C, Harris RA, Shores N, Epstein CB, Gjoneska E, Leung D, Xie W, Hawkins RD, Lister R, Hong C, Gascard P, Mungall AJ, Moore R, Chuah E, Tam A, Canfield TK, Hansen RS, Kaul R, Sabo PJ, Bansal MS, Carles A, Dixon JR, Farh KH, Feizi S, Karlic R, Kim AR, Kulkarni A, Li D, Lowdon R, Elliott G, Mercer TR, Neph SJ, Onuchic V, Polak P, Rajagopal N, Ray P, Sallari RC, Siebenthall KT, Sinnott-Armstrong NA, Stevens M, Thurman RE, Wu J, Zhang B, Zhou X, Beaudet AE, Boyer LA, De Jager PL, Farnham PJ, Fisher SJ, Haussler D, Jones SJ, Li W, Marra MA, McManus MT, Sunyaev S, Thomson JA, Tlsty TD, Tsai LH, Wang W, Waterland RA, Zhang MQ, Chadwick LH, Bernstein BE, Costello JF, Ecker JR, Hirst M, Meissner A, Milosavljevic A, Ren B, Stamatoyannopoulos JA, Wang T and Kellis M:

Integrative analysis of 111 reference human epigenomes. *Nature* 518: 317-330, 2015.

65. Li Q, Cheng Q, Chen Z, Peng R, Chen R, Ma Z, Wan X, Liu J, Meng M, Peng Z and Jiang B: MicroRNA-663 inhibits the proliferation, migration and invasion of glioblastoma cells via targeting TGF-beta1. *Oncology reports* 35: 1125-1134, 2016.

66. Yi C, Wang Q, Wang L, Huang Y, Li L, Liu L, Zhou X, Xie G, Kang T, Wang H, Zeng M, Ma J, Zeng Y and Yun JP: MiR-663, a microRNA targeting p21(WAF1/CIP1), promotes the proliferation and tumorigenesis of nasopharyngeal carcinoma. *Oncogene* 31: 4421-4433, 2012.

67. Cardenas H, Vieth E, Lee J, Segar M, Liu Y, Nephew KP and Matei D: TGF-beta induces global changes in DNA methylation during the epithelial-to-mesenchymal transition in ovarian cancer cells. *Epigenetics* 9: 1461-1472, 2014.

68. Suga Y, Sugai T, Uesugi N, Kawasaki T, Fukagawa T, Yamamoto E, Ishida K, Suzuki H and Sugiyama T: Molecular analysis of isolated tumor

glands from endometrial endometrioid adenocarcinomas. *Pathology international* 65: 240-249, 2015.

69. Phipps AI, Limburg PJ, Baron JA, Burnett-Hartman AN, Weisenberger DJ, Laird PW, Sinicrope FA, Rosty C, Buchanan DD, Potter JD and Newcomb PA: Association between molecular subtypes of colorectal cancer and patient survival. *Gastroenterology* 148: 77-87.e72, 2015.

70. Lister R, Pelizzola M, Dowen RH, Hawkins RD, Hon G, Tonti-Filippini J, Nery JR, Lee L, Ye Z, Ngo QM, Edsall L, Antosiewicz-Bourget J, Stewart R, Ruotti V, Millar AH, Thomson JA, Ren B and Ecker JR: Human DNA methylomes at base resolution show widespread epigenomic differences. *Nature* 462: 315-322, 2009.

71. Muraki Y, Banno K, Yanokura M, Kobayashi Y, Kawaguchi M, Nomura H, Hirasawa A, Susumu N and Aoki D: Epigenetic DNA hypermethylation: clinical applications in endometrial cancer (Review). *Oncology reports* 22: 967-972, 2009.

72. Zhang L, Wang W, Li X, He S, Yao J, Wang X, Zhang D and Sun X:

MicroRNA-155 promotes tumor growth of human hepatocellular carcinoma by targeting ARID2. International journal of oncology 2016.

73. Ma M, He M, Jiang Q, Yan Y, Guan S, Zhang J, Yu Z, Chen Q, Sun M, Yao W, Zhao H, Jin F and Wei M: MiR-487a Promotes TGF-beta1-induced EMT, the Migration and Invasion of Breast Cancer Cells by Directly Targeting MAGI2. International journal of biological sciences 12: 397-408, 2016.

74. Ge X, Liu X, Lin F, Li P, Liu K, Geng R, Dai C, Lin Y, Tang W, Wu Z, Chang J, Lu J and Li J: MicroRNA-421 regulated by HIF-1alpha promotes metastasis, inhibits apoptosis, and induces cisplatin resistance by targeting E-cadherin and caspase-3 in gastric cancer. Oncotarget 2016.

75. Feng S, Zhu X, Fan B, Xie D, Li T and Zhang X: miR19a3p targets PMEPA1 and induces prostate cancer cell proliferation, migration and invasion. Molecular medicine reports 2016.

76. Chen QY, Jiao DM, Wang J, Hu H, Tang X, Chen J, Mou H and Lu W: miR-206 regulates cisplatin resistance and EMT in human lung

adenocarcinoma cells partly by targeting MET. *Oncotarget* 2016.

77. Pan J, Hu H, Zhou Z, Sun L, Peng L, Yu L, Sun L, Liu J, Yang Z and Ran Y: Tumor-suppressive mir-663 gene induces mitotic catastrophe growth arrest in human gastric cancer cells. *Oncology reports* 24: 105-112, 2010.

78. Yan-Fang T, Jian N, Jun L, Na W, Pei-Fang X, Wen-Li Z, Dong W, Li P, Jian W, Xing F and Jian P: The promoter of miR-663 is hypermethylated in Chinese pediatric acute myeloid leukemia (AML). *BMC medical genetics* 14: 74, 2013.

79. Potapova A, Albat C, Hasemeier B, Haeussler K, Lamprecht S, Suerbaum S, Kreipe H and Lehmann U: Systematic cross-validation of 454 sequencing and pyrosequencing for the exact quantification of DNA methylation patterns with single CpG resolution. *BMC biotechnology* 11: 6, 2011.

80. Lehmann U, Hasemeier B, Christgen M, Muller M, Romermann D, Langer F and Kreipe H: Epigenetic inactivation of microRNA gene hsa-mir-9-1 in human breast cancer. *The Journal of pathology* 214: 17-24,

2008.

81. Yang Y, Wang LL, Li YH, Gao XN, Liu Y and Yu L: Effect of CpG island methylation on microRNA expression in the k-562 cell line. *Biochemical genetics* 50: 122-134, 2012.

82. Hong Q, Yu S, Geng X, Duan L, Zheng W, Fan M, Chen X and Wu D: High Concentrations of Uric Acid Inhibit Endothelial Cell Migration via miR-663 Which Regulates Phosphatase and Tensin Homolog by Targeting Transforming Growth Factor-beta1. *Microcirculation* (New York, N.Y. : 1994) 22: 306-314, 2015.

83. Narkiewicz J, Lapinska-Szumczyk S, Zurawa-Janicka D, Skorko-Glonek J, Emerich J and Lipinska B: Expression of human HtrA1, HtrA2, HtrA3 and TGF-beta1 genes in primary endometrial cancer. *Oncology reports* 21: 1529-1537, 2009.

84. Kogure T, Kondo Y, Kakazu E, Ninomiya M, Kimura O and Shimosegawa T: Involvement of miRNA-29a in epigenetic regulation of transforming growth factor-beta-induced epithelial-mesenchymal transition

in hepatocellular carcinoma. *Hepatology research : the official journal of the Japan Society of Hepatology* 44: 907-919, 2014.

85. Zhang Q, Chen L, Helfand BT, Jang TL, Sharma V, Kozlowski J, Kuzel TM, Zhu LJ, Yang XJ, Javonovic B, Guo Y, Lonning S, Harper J, Teicher BA, Brendler C, Yu N, Catalona WJ and Lee C: TGF-beta regulates DNA methyltransferase expression in prostate cancer, correlates with aggressive capabilities, and predicts disease recurrence. *PloS one* 6: e25168, 2011.

86. Nosho K, Shima K, Irahara N, Kure S, Baba Y, Kirkner GJ, Chen L, Gokhale S, Hazra A, Spiegelman D, Giovannucci EL, Jaenisch R, Fuchs CS and Ogino S: DNMT3B expression might contribute to CpG island methylator phenotype in colorectal cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research* 15: 3663-3671, 2009.

87. Roll JD, Rivenbark AG, Jones WD and Coleman WB: DNMT3b overexpression contributes to a hypermethylator phenotype in human breast

cancer cell lines. *Mol Cancer* 7: 15, 2008.

88. Kanai Y, Ushijima S, Kondo Y, Nakanishi Y and Hirohashi S: DNA methyltransferase expression and DNA methylation of CPG islands and peri-centromeric satellite regions in human colorectal and stomach cancers. *International journal of cancer* 91: 205-212, 2001.

89. Samowitz WS, Curtin K, Lin HH, Robertson MA, Schaffer D, Nichols M, Gruenthal K, Leppert MF and Slattery ML: The colon cancer burden of genetically defined hereditary nonpolyposis colon cancer. *Gastroenterology* 121: 830-838, 2001.

90. Ollikainen M, Abdel-Rahman WM, Moisio AL, Lindroos A, Kariola R, Jarvela I, Poyhonen M, Butzow R and Peltomaki P: Molecular analysis of familial endometrial carcinoma: a manifestation of hereditary nonpolyposis colorectal cancer or a separate syndrome? *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 23: 4609-4616, 2005.

91. Ollikainen M, Hannelius U, Lindgren CM, Abdel-Rahman WM,



Kere J and Peltomaki P: Mechanisms of inactivation of MLH1 in hereditary nonpolyposis colorectal carcinoma: a novel approach. *Oncogene* 26: 4541-4549, 2007.

92. Lao VV and Grady WM: Epigenetics and colorectal cancer. *Nature reviews. Gastroenterology & hepatology* 8: 686-700, 2011.

93. Samowitz WS, Albertsen H, Sweeney C, Herrick J, Caan BJ, Anderson KE, Wolff RK and Slattery ML: Association of smoking, CpG island methylator phenotype, and V600E BRAF mutations in colon cancer. *Journal of the National Cancer Institute* 98: 1731-1738, 2006.

94. Sanchez JA, DeJulius KL, Bronner M, Church JM and Kalady MF: Relative role of methylator and tumor suppressor pathways in ulcerative colitis-associated colon cancer. *Inflammatory bowel diseases* 17: 1966-1970, 2011.

95. Olaru AV, Cheng Y, Agarwal R, Yang J, David S, Abraham JM, Yu W, Kwon JH, Lazarev M, Brant SR, Marohn MR, Hutcheon DF, Harpaz N, Meltzer SJ and Mori Y: Unique patterns of CpG island methylation in

inflammatory bowel disease-associated colorectal cancers. *Inflammatory bowel diseases* 18: 641-648, 2012.



## Acknowledgments

I would like to express my gratitude to all those who gave me the opportunity to complete this thesis, which would not have been possible without their support.

First of all, I would like to express my deepest appreciation to Professor Kuniya Abe for all his guidance and support throughout the course of this study. His knowledge and advice were invaluable to me.

Next, I wish to thank Dr. M. Adachi (Keio University, Japan) for sample collection and clinical data analysis, Drs. K. Banno and D. Aoki (Keio University, Japan) for diagnosis and treatment of Lynch syndrome. I also thank all the member of our laboratory.

Finally, I am really grateful to my father (Dr. Minoru Y.), mother (Dr. Mieko Y.), husband (Mr. Tomoshi M.) and son (Jun M.) for their love and support.

*Megumi Moroyama*

*To the memory of my late father.*